**Encephalitozoon cuniculi**

Infection in Rabbits

Carly N. Jordan, BS, MS
Anne M. Zajac, DVM, PhD
David S. Lindsay, PhD
Virginia–Maryland Regional College of Veterinary Medicine

**ABSTRACT:** *Encephalitozoon cuniculi* is an important protozoan parasite of pet rabbits. It most commonly infects the brain and kidneys, causing clinical signs such as head tilt, ataxia, and hindlimb paralysis, although most cases of infection are subclinical. Transmission of this parasite primarily occurs transplacentally or by ingestion or inhalation of spores passed in the urine. Although no prevalence data exist for infection in rabbits in the United States, serologic studies indicate that exposure is common among rabbits throughout Europe and Africa.

*Encephalitozoon cuniculi* is an obligate intracellular parasite in the phylum Microspora, which is defined by its resistant spore stage. Microsporidia receive little attention in veterinary medical curricula because they are considered important only as parasites of rabbits and fish. Microsporidia are important parasites of humans as well, with some species causing severe disease and death in immunocompromised individuals. Microsporidia are among the smallest protozoa known and have the smallest genome (i.e., 2.9 million base pairs) of all eukaryotes. Microsporidia have many characteristics in common with prokaryotes. For example, they lack mitochondria, centrioles, and peroxisomes. In addition, the Golgi apparatus is atypical and primitive, and ribosomes of Microsporidia closely resemble those of prokaryotes. Sequencing of microsporidial genes has suggested a relationship with fungi. This theory is supported by the presence of chitin in the spore wall and by intranuclear division.

**LIFE CYCLE OF ENCEPHALITOZOON CUNICULI**

The life cycle of *E. cuniculi* takes 3 to 5 weeks to complete. Hosts are infected by ingestion or inhalation of spores or by transplacental transmission (Figure 1). The spore penetrates a host cell via the polar tube. The polaroplast swells, and the anchoring disk stabilizes the tube. The polar tube is inverted while extruded, and membranes from the polaroplast connect with the tube to extend its length. The extrusion occurs with great force and allows penetration of the host cell without disturbance of the host cell membrane. The posterior vacuole swells and forces the sporoplasm and nucleus through the polar tube (Figure 2). The tip of the polar tube inside the host cell envelopes the sporoplasm to become the parasite’s outer membrane.

The sporoplasm divides, creating meronts, the major proliferative stage that divides by binary fission. This stage exists in a parasitophorous vacuole (PV) lined by the host cell membrane. Meronts may undergo several replications before converting into sporonts. In most species, the sporonts exist within a PV.
Sporonts convert to sporoblasts, either immediately or after one division. Sporoblast is the stage that synthesizes the polar tube and its accessory organelles to become mature spores. After one host cell is full of mature spores, the cell bursts, and the spores are released (Figure 3). These spores are resistant to environmental factors and can remain viable for several years. Spores are passed in the urine of rabbits, beginning around 35 days after infection, and continue to be excreted for 2 to 3 months.

**ENCEPHALITOZOOONOSIS**

*E. cuniculi* was first identified in 1922 in a colony of research rabbits and is now recognized as a common pathogen infecting pet rabbits. There have been no large-scale studies of encephalitozoonosis in rabbits in the United States, but surveys conducted in Europe have shown high rates of infection in asymptomatic rabbits (7% to 42%) and those with neurologic signs (40% to 85%; Table 1).

Adult rabbits are infected by ingestion or inhalation of spores passed in the urine, but vertical transmission can also occur. Transplacental transmission has been confirmed by detection of *E. cuniculi* DNA in placentas and fetuses delivered by cesarean section. Transplacental transmission can eliminate the pathogen in a breeding colony. The infectious dose required to cause disease in 50% of rabbits is only 46 *E. cuniculi* spores.

The route of *E. cuniculi* infection in rabbits begins with invasion of the intestinal epithelium, where parasites undergo several rounds of replication. From there, infective spores are disseminated throughout the body—first to the heart, lungs, liver, and spleen. To this point, rabbits are usually asymptomatic, and damage to these organs is limited. The final destinations of *E. cuniculi* in rabbits are the kidneys, eyes, and brain, where the parasites (7% to 42%) and those with neurologic signs (40% to 85%; Table 1).

Figure 1. Life cycle of *E. cuniculi*. The cellular details are visible only by using transmission electron microscopy.
site reaches equilibrium with the host’s immune response. The parasite may reside in these organs indefinitely without ever causing clinical signs in an animal. However, some rabbits are more affected by invasion of these organs, and clinical disease occurs. Rabbits younger than 6 weeks of age are more likely to develop severe disease.

**Neurologic Disease**

Most *E. cuniculi* infections are asymptomatic, but when clinical signs are observed, they are usually neurologic. The onset of disease in rabbits can be sudden, and severe cases may be rapidly fatal. Early signs of disease include vestibular disease, head tilt, and ataxia. Hindlimb weakness or paralysis is common with loss of postural reactions and sluggish spinal reflexes. Head tremors and seizures may occur because of an inflammatory response to ruptured brain cells. Seizures typically occur suddenly, and some rabbits are left blind or comatose, whereas others recover completely. Rabbits chronically affected by *E. cuniculi* may demonstrate stargazing, loss of balance, and swaying or nodding when at rest. Animals may become aggressive and lose sight or hearing.

Examination of brain tissue typically demonstrates meningoencephalitis with perivascular cuffing, and focal granulomas of glial cells and lymphocytes may also be observed. Multifocal mineralization is also observed in some cases. Brain lesions are usually not observed for at least 8 weeks after antibodies are first detected or approximately 3 months after infection.

**Kidney Disease**

Infection of the kidneys may lead to polydipsia and polyuria as well as urinary incontinence, although it is not clear whether this is due to neurologic or kidney disease. Gross examination of the kidneys reveals a characteristic pitted appearance with irregular grayish white spots and depressions. The kidneys are shrunken, fibrotic, and pale. Histologic examination reveals granulomatous interstitial nephritis with infiltration by macrophages. Degeneration of renal tubules is also observed.

Blood biochemistry may show a mild elevation in blood urea, with levels around 150 mg/dl. Creatinine concentrations can reach 6 mg/dl, and serum potassium may also be elevated. Some animals experience anemia and low hemoglobin and erythrocyte counts.

**Ocular Disease**

Ocular disease is common in *E. cuniculi*–infected rabbits. Some researchers believe that *E. cuniculi* spores infect the developing lens in utero, when the lens capsule is thin or absent. Infection of the lens results in phacoclastic uveitis, another condition that may also be caused by pasteurellosis. The anterior lens capsule ruptures, leading to zonal granulomatous lens uveitis, with inflammation occurring near the capsule break. Severe inflammation of the globe can lead to loss of vision. Cataracts may occur as a result of disruption of the lens fibers, although vision is usually retained.

**DIFFERENTIAL DIAGNOSIS**

Presentation of encephalitozoonosis in rabbits may be easily confused with disease caused by several other...
encephalitozoonosis, although the most common clinical signs associated with P multocida are upper respiratory tract infection and pneumonia. E cuniculi infection causes central vestibular disease in rabbits, and P multocida infection typically causes peripheral vestibular disease. However, P multocida infection can also result in central lesions if abscesses form along the vestibular tract.

Hematology can be useful in differentiating between E cuniculi and P multocida infections. No hematologic changes are associated with E cuniculi infection, but patients with pasteurellosis often have neutrophilia and left shift. Radiography of the head may show changes in the tympanic bullae associated with chronic otitis media due to P multocida infection. Serology is useful only for ruling out pathogens, as a high percentage of rabbits have been exposed to one of these pathogens.

METHODS OF DIAGNOSIS
Serology
Serologic testing is the most common method of diagnosing E cuniculi infection. In naturally infected rabbits, antibodies to E cuniculi are usually detectable by 4 weeks after infection, with peak titers observed around 9 weeks. The indirect immunofluorescent antibody (IFA) test is the gold standard for serologic diagnosis of Microsporidia infection, and IFA titers greater than 1:20 are considered positive. Monoclonal and polyclonal antibodies have been developed for E cuniculi infection for use in IFA tests on an experimental basis and in some diagnostic laboratories.

ELISA is another method of serologically diagnosing Microsporidia infection. Two groups of researchers have evaluated ELISA for diagnosing E cuniculi infection by comparing the results with IFA. One group examined 135 rabbits diagnosed with infection by histology and 75 rabbits without a history of illness. A few samples were positive by only one method, but no statistically significant differences were calculated between the sensitivities and specificities of the two assays. Another group used a modified ELISA (dot-ELISA) to detect antibodies to E cuniculi in rabbits, dogs, squirrel monkeys, and mice. Comparison of the dot-ELISA with IFA showed agreement between the two assays.

Table 1. Prevalence of E cuniculi in Rabbits as Determined by Serologic Surveys

<table>
<thead>
<tr>
<th>Location</th>
<th>Health</th>
<th>No. Tested</th>
<th>No. Positive</th>
<th>% Positive</th>
<th>Study</th>
</tr>
</thead>
<tbody>
<tr>
<td>Slovakia</td>
<td>No information available</td>
<td>571</td>
<td>238</td>
<td>41.7%</td>
<td>Halanová et al</td>
</tr>
<tr>
<td>Switzerland</td>
<td>Asymptomatic</td>
<td>292</td>
<td>22</td>
<td>7.5%</td>
<td>Deplazes et al</td>
</tr>
<tr>
<td></td>
<td>Neurologic signs</td>
<td>72</td>
<td>61</td>
<td>84.7%</td>
<td>Deplazes et al</td>
</tr>
<tr>
<td>W. Yorkshire, UK</td>
<td>Asymptomatic</td>
<td>38</td>
<td>14</td>
<td>36.8%</td>
<td>Harcourt-Brown et al</td>
</tr>
<tr>
<td></td>
<td>Neurologic signs</td>
<td>58</td>
<td>23</td>
<td>39.7%</td>
<td>Harcourt-Brown et al</td>
</tr>
<tr>
<td></td>
<td>Ocular lesions</td>
<td>8</td>
<td>8</td>
<td>100%</td>
<td>Harcourt-Brown et al</td>
</tr>
<tr>
<td>Spain</td>
<td>No information available</td>
<td>22</td>
<td>2</td>
<td>9.1%</td>
<td>Lores et al</td>
</tr>
</tbody>
</table>

Serologic surveys in Europe report the prevalence of E cuniculi in rabbits with neurologic disease to be 40% to 85%.
Light Microscopy

Examination of tissue samples or secretions is another common method of diagnosing *E. cuniculi* infection in rabbits. Kidney biopsy can reveal spores approximately 5 weeks after antibodies are first produced or approximately 9 weeks after infection. In infected patients with uveitis, *E. cuniculi* spores can be found in the liquefied lens cortex. Several staining techniques are available for identifying Microsporidia spores; however, current staining methods can, at best, identify Microsporidia but cannot distinguish between *Encephalitozoon* spp. These methods are also hindered by the fact that Microsporidia are often mistaken for bacteria or yeasts.

Calcofluor white M2R is a chemofluorescent stain commonly used to identify Microsporidia in urine sediments, fecal smears, and respiratory lavages. This is the most sensitive method of staining Microsporidia, and the procedure can be completed in only 15 minutes. However, this stain is not specific for Microsporidia, and spores may be confused with yeasts or other particulate matter in the sample. Because a compound microscope with an ultraviolet light source is needed for this staining method, it is used only in diagnostic laboratories.

Chromotrope-based (modified trichrome) staining methods are also used to identify Microsporidia in feces, urine sediment, or respiratory secretion samples. Chromotrope stains color spores pinkish-red, and quick-hot Gram’s–chromotrope stains color spores dark violet. Characteristic morphologic features of Microsporidia that are visible using these stains are a belt-like stripe, a vacuole, and gram-positive granules. Spores may be confused with bacteria or yeasts.

Giemsa and Gram’s stains are not recommended for fecal smears because they do not differentiate between Microsporidia and bacteria or yeast that may be present in the sample. However, Giemsa stains may be useful in urine or tissue samples, and spores stain light blue. Microsporidia spores are gram-positive in gram-stained tissue sections (Figure 4), which can aid in identification in histologic samples.

Electron Microscopy

Transmission electron microscopy, although impractical for most veterinarians, can be used to differentiate among the four species of Microsporidia that most commonly infect humans and companion animals: *E. cuniculi*, *Encephalitozoon intestinalis*, *Encephalitozoon hellem*, and *Enterocytozoon bieneusi*. *Ent. bieneusi* is slightly smaller than *Encephalitozoon* spp: *Ent. bieneusi* is 0.5 × 1.5 µm, whereas *Encephalitozoon* spp range from 1 to 1.5 × 2 to 2.5 µm. The number of polar tube coils is useful in differentiating between *Encephalitozoon* spp (Table 2). *E. intestinalis* can be distinguished from all other *Encephalitozoon* spp by examination of the PV within an infected host cell. All *Encephalitozoon* species develop within a PV, but *E. intestinalis* is the only species of Microsporidia known to exist in a septated PV that isolates each developing spore (Figure 5). *Ent. bieneusi* does not create a PV; instead, it develops in direct contact with the host cell cytoplasm.

Clinical signs of *E. cuniculi* infection in rabbits are primarily neurologic, including head tilt, paralysis, and seizures.
Polymerase chain reaction (PCR) testing can be used to positively identify species of Microsporida from feces, urine, or tissue samples. Specific primers are available for *E. cuniculi*, *E. intestinalis*, and *E. hellem* that amplify diagnostic fragments of 549, 520, and 546 base pairs, respectively. This method is a definitive way to identify Microsporida at the species level, but PCR testing is available only in select research laboratories.

**TREATMENT**

Very little information is available on treating microsporidiosis in animals. Albendazole is the most common drug used to treat human Microsporida infections. Albendazole acts by interfering with the polymerization of β-tubulin, thereby preventing nuclear division. In vitro models have shown that albendazole is more than 90% effective in inhibiting the growth of *E. cuniculi*. No drugs have been approved to treat *E. cuniculi* infection in rabbits. However, treating rabbits with albendazole (15 mg/kg PO q24h) can reduce clinical signs of disease and help stop passage of spores in urine.

One case report describes the use of oral albendazole together with topical prednisolone in the successful treatment of a rabbit with phacoclastic uveitis. Phacoemulsification of the lens was performed after the rabbit did not respond well to initial treatment with 1% topical prednisolone, but 4 months after surgery, the rabbit presented with keratitis and a large inflammatory mass. The rabbit was treated with albendazole (30 mg/kg PO) for 4 weeks, and then the dose was reduced to 15 mg/kg PO for an additional 4 weeks. Prednisolone (1%) was also administered as often as every 6 hours. After 8 weeks of treatment, the inflammation resolved but the cornea was slightly scarred.

A related drug, fenbendazole, has been shown to prevent and treat *E. cuniculi* infection in rabbits. Eight naturally infected rabbits were treated with fenbendazole for 4 weeks; afterward, no parasites could be isolated from brain tissue. One group of four rabbits was administered fenbendazole (0.1 ml/kg PO bid for 7 days) in suspension before experimental inoculation.

**Molecular Techniques**

Polymerase chain reaction (PCR) testing can be used to positively identify species of Microsporida from feces, urine, or tissue samples. Specific primers are available for *E. cuniculi*, *E. intestinalis*, and *E. hellem* that amplify diagnostic fragments of 549, 520, and 546 base pairs, respectively. This method is a definitive way to identify Microsporida at the species level, but PCR testing is available only in select research laboratories.

**Table 2. Morphologic Characteristics of Common Microsporida**

<table>
<thead>
<tr>
<th></th>
<th><em>E. cuniculi</em></th>
<th><em>E. intestinalis</em></th>
<th><em>E. hellem</em></th>
<th>Ent. bieneusi</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spore dimensions (µm)</td>
<td>1.5 × 2.5</td>
<td>1.2 × 2</td>
<td>1 × 2.5</td>
<td>0.5 × 1.5</td>
</tr>
<tr>
<td>No. of polar tube coils (single row)</td>
<td>5–7</td>
<td>4–7</td>
<td>4–9</td>
<td>4–5 (double row)</td>
</tr>
<tr>
<td>Development in host cell</td>
<td>PV</td>
<td>Septated PV</td>
<td>PV</td>
<td>Direct contact with cytoplasm</td>
</tr>
</tbody>
</table>

*PV* = parasitophorous vacuole.
with *E. cuniculi*, and treatment continued for 21 days after inoculation. The other group of four rabbits was fed 20 mg/kg of fenbendazole in pellet form from 7 days before to 2 days after infection. Both treatments were sufficient to prevent infection. The rabbits did not seroconvert, and spores were not isolated from the brain tissue of rabbits from either group when they were examined 120 days after infection.

Anecdotal evidence suggests that tetracyclines may be administered to suppress *E. cuniculi*, although they are not sufficient to clear infections. They can be given in water at a concentration of 500 mg/L or as oral doses of 20 mg/kg bid. In mild cases, clinical signs can be treated using steroids to control inflammation. Corticosteroids are most effective for treating uveitis, and dexamethasone can be given at a dose of 2 mg/kg SC or IM. However, corticosteroid administration may enhance parasite infection, so animals should be closely monitored.

**CONCLUSION**

*E. cuniculi* is an important pathogen of rabbits, and serologic surveys suggest that infection is common throughout Europe and Africa. Clinical signs usually result from infection of the brain and include ataxia, hindlimb paralysis, and seizures. Infection of the kidneys and eyes is also common, and clinical signs of ocular infection may include uveitis and cataracts. Transmission of the parasite occurs transplacentally or by ingestion or inhalation of spores. Serum antibodies to *E. cuniculi* may be identified using immunologic assays such as ELISA and IFA. Samples from urine, feces, and tissue can be tested via PCR testing for the presence of *E. cuniculi* DNA. No drugs are currently licensed for treating microsporidial infection in rabbits, but several reports suggest that human treatments are effective in controlling clinical signs and reducing parasite shedding. Although no direct evidence of animal-to-human transmission exists, it is widely accepted that *E. cuniculi* is a zoonotic parasite, and immunocompromised pet owners should be made aware of the risks.

**REFERENCES**


ARTICLE #1 CE TEST

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1. Transmission of E. cuniculi infection in rabbits occurs by
   a. ingestion of spores.
   b. inhalation of spores.
   c. transplacental transmission.
   d. all of the above

2. Which is the major proliferative stage of Microsporidia?
   a. sporule
   b. meront
   c. sporont
   d. sporoblast

3. Antibodies to E. cuniculi infection are usually detectable by ______ weeks after infection, and spores are shed in urine by ______ days after infection.
   a. 4; 21
   b. 4; 35
   c. 5; 21
   d. 5; 35

4. The ______ is(are) not commonly infected by E. cuniculi.
   a. brain
   b. liver
   c. kidneys
   d. eyes

5. The infectious dose required for initiating E. cuniculi infection in rabbits is ______ spores.
   a. 12
   b. 23
   c. 46
   d. 64

6. Which clinical sign is not typically observed during the acute stage of infection in rabbits?
   a. head tilt
   b. ataxia
   c. vestibular disease
   d. aggressive behavior

7. Which pathogen(s) should be included in the differential diagnosis of E. cuniculi infection?
   a. T. gondii
   b. P. multocida
   c. Eimeria stiedae
   d. a and b

8. What is the gold standard for diagnosis of E. cuniculi infection?
   a. Gram’s stain
   b. IFA testing
   c. ELISA
   d. PCR testing

9. Spores of E. cuniculi develop
   a. in direct contact with the host-cell cytoplasm, together with other spores.
   b. in direct contact with the host-cell cytoplasm, isolated from one another.
   c. within a PV, together with other spores.
   d. within a PV, isolated from one another.

10. Which drug is approved for treating E. cuniculi infection in rabbits?
    a. albendazole
    b. dexamethasone
    c. fenbendazole
    d. none of the above